



Can Ammonium Stress Be Positive for Plant Performance?

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INTRODUCTION

Ammonium ($\text{NH}_3/\text{NH}_4^+$) nutrition is considered as a universal stressful situation (recently reviewed in Li et al., 2014; Esteban et al., 2016; Liu and Von Wirén, 2017). Briefly, the most common symptom of ammonium nutrition is reduced biomass accumulation with respect to non-stressed plants. Growth inhibition has been associated with the high energy cost to control $\text{NH}_3/\text{NH}_4^+$ level in tissues. Among others, ammonium stress has been related with deregulation of pH homeostasis, ion imbalance, impaired nitrate signaling, or hormone deregulation (Li et al., 2014; Esteban et al., 2016; Liu and Von Wirén, 2017). Although ammonium stress affects virtually every plant species, the degree of stress it generates is variable and high intraspecific and interspecific variability towards ammonium nutrition has been reported. Some species/genotypes display ammonium preference, while others show extreme sensitivity when growing with ammonium. Regarding the response of a certain genotype, as for almost every stress, there exists a continuum in the response upon ammonium nutrition, which mostly depends on the concentration of NH_4^+ in the root medium. Overall, ammonium tolerance could be defined as a situation where the plant is somehow sensing and responding towards ammonium stress prior to suffering a serious damage such as chlorosis or cell death. Sole ammonium nutrition is an artificial situation that only takes place when growing plants without soil, either in laboratory conditions or for example when growing crops in pure hydroponics or in inert substrates such as rockwool or perlite. In agricultural fields, exclusive ammonium nutrition does not exist; however, the use of nitrification inhibitors together with ammonium fertilizers or organic fertilizers makes ammonium stable and at high concentrations in the soil for several weeks. From a farmer's point of view, a potential moderate reduction in yield caused by ammonium stress could be compensated with benefits such as an increase in the resistance of the crop against biotic or abiotic constraints and also with obtaining of products of higher quality (**Figure 1**). Moreover, the use of ammonium-based fertilizers together with inhibitors of nitrification has been extensively shown to mitigate the impact of nitrogen fertilizers on the environment (Sanz-Cobena et al., 2017). Although sophisticated management would be needed, avoiding ammonium stress could be reached by, for instance, fertigation or frequent additions of small amounts of ammonium-based fertilizers in water delivered through micro-irrigation.

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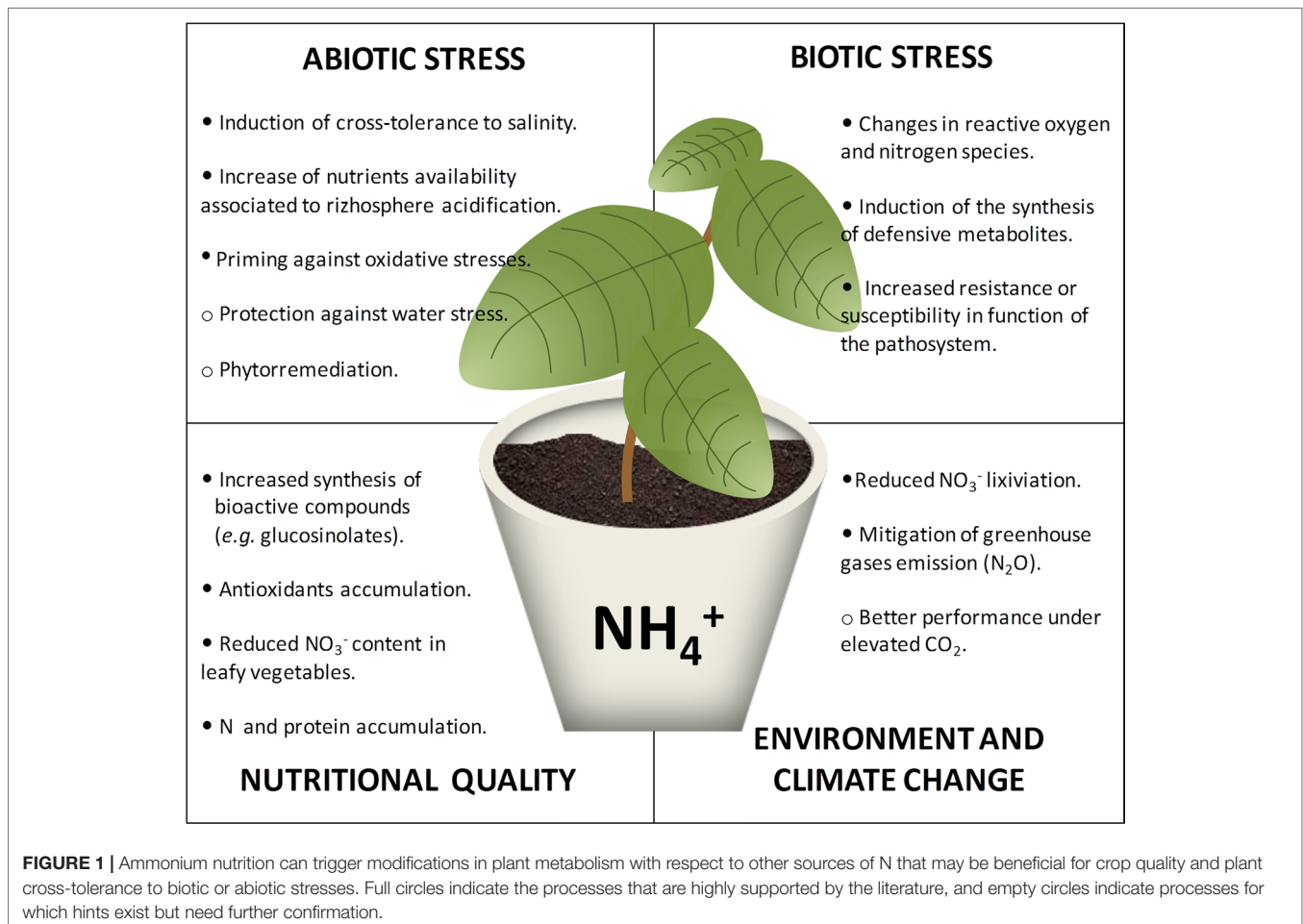
AMMONIUM NUTRITION MAY IMPROVE THE QUALITY OF CROPS

The main cause of ammonium toxicity is probably the over-accumulation of free NH_4^+ in the cytosol and the problems derived from cell efforts to get rid of it. The cell has several logical strategies to keep NH_4^+ levels under control: 1) NH_4^+ efflux to the apoplast/rhizosphere, 2) NH_4^+ storage in the vacuole, and 3) NH_4^+ assimilation into organic compounds.

In line with the third strategy to avoid excessive cytosolic NH_4^+ accumulation, the induction of the synthesis of N-reduced compounds is a classical plant response to ammonium nutrition, and indeed, the accumulation of total free amino acids can be considered as a marker of ammonium stress (Sarasketa et al., 2014). In general terms, crop quality is associated with the protein content of food products, notably in grains, which is dependent on the crops' capacity to efficiently use the available nitrogen. In addition, the nutritional value and/or quality of food is associated with its content in minerals and in health-promoting secondary metabolites such as antioxidants. In this line of evidence, several works have reported an improvement of the nutritional quality of a number of crops when they are grown under ammonium nutrition. A higher protein accumulation is common in plants grown with NH_4^+ supply, and for instance, a positive effect of ammonium nutrition

with respect to nitrate (NO_3^-) was reported in the protein content of wheat grain and in the gliadins/glutenins ratio, overall increasing wheat bread-making quality (Fuertes-Mendizábal et al., 2013).

In Brassicaceae, glucosinolates (GLS) represent an abundant family of secondary metabolites derived from amino acids. GLS degradation products participate in cruciferous plant defense against herbivores. Moreover, they are responsible for the characteristic flavor of the cruciferous vegetables. Importantly, certain GLS breakdown products possess health-protective capacities, particularly anticarcinogenic activity, and hence, GLS content is associated with cruciferous nutritional quality. Currently, big efforts are being dedicated to manipulate GLS levels in order to produce new and improved commercial cruciferous crop varieties (Traka et al., 2013). Regarding ammonium-based nutrition, recent studies have reported that the synthesis of GLS is stimulated in leaves of plants grown with NH_4^+ as N source, such as in broccoli, oilseed rape, and Chinese kale (La et al., 2013; Marino et al., 2016; Coletto et al., 2017). Notably, glucoraphanin content, whose degradation yields sulforaphane, the most promising and characterized anticancer isothiocyanate, increases in ammonium-fed broccoli and oilseed rape (Marino et al., 2016; Coletto et al., 2017). Whether GLS accumulation is just a consequence of ammonium assimilation increase or whether they possess a regulatory role during ammonium stress is a question for further elucidation.



Another aspect of crop quality is the control of NO_3^- accumulation in plant edible parts, notably in leafy vegetables such as spinach or lettuce. This is a subject of concern because it can turn to nitric compounds, which have been linked to increased risk of cancer and methemoglobinemia (Umar and Iqbal, 2007). Accordingly, growing plants with increased amounts of NH_4^+ with respect to NO_3^- clearly reduces the quantity of NO_3^- accumulated in plant tissues and thus its associated risks (Santamaria et al., 2001; Irigoyen et al., 2006).

AMMONIUM NUTRITION MAY PROTECT PLANTS FROM PATHOGEN ATTACK

Nitrogen metabolism is closely connected to plant immunity. Among others, it provides the necessary building blocks to synthesize most of the defense-related secondary metabolites and is central for NO production whose role in plant–pathogen interaction has been widely reported (Santana et al., 2017). Nitrogen source has been shown to have an impact on plant immunity. A number of studies have reported that plants grown with NO_3^- displayed increased resistance to pathogen attack with respect to plants grown with NH_4^+ ; for instance, in tobacco exposed to *Pseudomonas syringae* (Gupta et al., 2013), cucumber infected with *Fusarium oxysporum* (Wang et al., 2016), or rice attacked by *Rhizoctonia solani* (Chi et al., 2019). This higher resistance has been associated with higher NO production in nitrate-fed plants, hormone signaling, or decreased citrate exudation, among others (Gupta et al., 2013; Mure et al., 2016; Wang et al., 2016). In contrast, several works have reported increased resistance in ammonium-fed plants such as tomato exposed to *P. syringae* (Fernández-Crespo et al., 2015) or to *F. oxysporum* (Lopez-Berges et al., 2010) and potato facing *Verticillium* wilt (Huber, 1989). In this case, the beneficial priming effect of NH_4^+ has been related to an increased reactive oxygen species burst and polyamine synthesis in ammonium-fed plants (Fernández-Crespo et al., 2015). Moreover, transcriptomic analyses have reported that ammonium induces the upregulation of genes associated with plant defense and immunity (Patterson et al., 2010; Vega-Mas et al., 2017). Interestingly, the overexpression of rice ammonium transporter *AMT1;2* conferred resistance against *R. solani* (Chi et al., 2019). In contrast, Arabidopsis *amt1.1* knockout mutant exhibited less disease symptoms than did wild-type plants infected with *P. syringae* and *Plectosphaerella cucumerina* (Pastor et al., 2014).

In another line of evidence, the above-reported increase in GLS synthesis might be also increasing the resistance of cruciferous plants notably against herbivores (Marino et al., 2016). Similarly, the stimulation of the synthesis of γ -aminobutyric acid (GABA) is also frequent under ammonium nutrition, for instance, in tobacco plants (Gupta et al., 2013). GABA is a signal molecule common to animals and plants. Its accumulation reveals a stress-specific pattern consistent with a physiological response leading to stress mitigation and is also involved in plant response to pathogens (Kinnarsley and Turano, 2000; Bown and Shelp, 2016). GABA accumulation appeared detrimental for plant defense (Gupta et al., 2013); nevertheless, further experimentation is needed to fully

decipher the role of GABA in the connection between N-source use and plant immunity. Overall, the interaction between NH_4^+ and plant defense is clear, but the potential benefit of ammonium stress would be dependent on the plant pathosystem, and therefore, no general rule can be drawn.

AMMONIUM NUTRITION MAY IMPROVE THE CROSS-TOLERANCE TO OTHER ABIOTIC STRESSES

A number of the responses that ammonium nutrition may trigger are defensive mechanisms that are common to different abiotic stress situations. Interestingly, the onset of these mechanisms may prevent damage from other simultaneous or subsequent stresses. Salinity is one of the most detrimental abiotic stresses, and the type of N nutrition differentially affects plants living under high salt contents. For example, the C4 halophyte *Spartina alterniflora* displayed improved performance when grown with NH_4^+ as N source, and NH_4^+ benefits were associated with higher antioxidant enzyme activities (Hessini et al., 2013). Intriguingly, although antioxidant machinery induction was higher, *S. alterniflora* NH_4^+ preference was lost under drought (Hessini et al., 2017). While *S. alterniflora* is a highly tolerant plant to ammonium nutrition, similar positive effect can be observed in other species. For instance, in the citrus *citrange* Carrizo, NH_4^+ nutrition promoted its resistance to salinity conditions, inducing, among other responses, lower Cl^- uptake. The mechanisms of action again showed that plant antioxidant machinery, notably glutathione metabolism, was part of a common NH_4^+ response that primed resistance to subsequent salt stress (Fernández-Crespo et al., 2014). Similarly, NH_4^+ -induced cross-acclimation to salinity stress has also been reported in *Sorghum bicolor* (de Souza Miranda et al., 2017). Ammonium nutrition improved K^+/Na^+ homeostasis notably by reducing Na^+ loading into the xylem in agreement with the observed higher proton pumps and Salt Overlay Sensitive 1 (SOS1) Na^+/H^+ antiporter activity. In general, ammonium acted as an efficient signal to activate responses involved in the regulation of Na^+ homeostasis, leading to salt tolerance in sorghum plants (de Souza Miranda et al., 2017). More recently, the benefit of NH_4^+ as a primer of resistance to salinity has also been reported in maize (Hessini et al., 2019).

Previous ammonium nutrition has also been shown to ameliorate water stress resistance. Thus, Gao et al. (2010) showed an important fresh weight increase in rice plants under polyethylene glycol (PEG)-induced water stress when ammonium nutrition was the source of N, while either nitrate or mixed sources significantly decreased fresh weight under water stress. This effect was suggested to be related to higher aquaporin activity, which takes place in ammonium-grown plants, independently of the water stress, and which relates to a better usage of water under NH_4^+ nutrition (Gao et al., 2010). Similarly, the alleviation of PEG-induced water stress in ammonium-fed rice seedlings has been related with sustained NH_4^+ uptake and assimilation (Cao et al., 2018). Indeed, it has been suggested that increasing nitrogen uptake and assimilation, among others in

tomato (Sánchez-Rodríguez et al., 2011) and in *Malus prunifolia* (Huang et al., 2018), could increase the cell osmotic adjustment capacity to protect plants against water stress.

Ammonium uptake is known to involve proton extrusion to the apoplast/rhizosphere. Rhizosphere acidification is often deleterious for plant growth (Shavrukov and Hirai, 2016), and ammonium stress symptoms usually increases at more acidic pHs (Chaillou et al., 1991; Sarasketa et al., 2016). However, notably, in neutral/alkaline soils, ammonium nutrition may increase the availability of certain nutrients, such as iron or phosphorus, and improve plant growth (Gahoonia et al., 1992; Logan et al., 2000). Among others, the increase in nutrient availability induced by pH acidification has also been put forward as one of the reasons that may confer pathogen resistance to plants grown under ammonium nutrition (Leusch and Buchenauer, 1988; Huber and McCay-Buis, 1993). Furthermore, the combination of ammonium fertilization with plant-growth-promoting microorganisms may have a positive synergistic effect on plant performance (Bradáčová et al., 2019; Mpanga et al., 2019).

Ammonium nutrition has also been talked about in relation to its interaction with plant response to elevated atmospheric CO₂ due to the hypothesis of Bloom et al. (2010), stating that C3 plants respond more positively to elevated CO₂ under ammonium nutrition than under nitrate nutrition. It is suggested that elevated CO₂ inhibits the plant photoreduction of NO₃⁻ and consequently reduces total plant N assimilation and growth (Rubio-Asensio and Bloom, 2017). However, this hypothesis is today under great debate, and a number of works do not support it (Vega-Mas et al., 2015; Andrews et al., 2018). On the whole, the magnitude of the challenge that climate change adaptation implies for agriculture deserves further research to discard or confirm the potential benefit of ammonium nutrition for plant performance.

Beyond drought and salinity, ammonium nutrition has also been suggested to contribute to other stressful situations such as the tolerance of cucumber to phenanthrene, a persistent

polycyclic aromatic hydrocarbon commonly found in soil and sediments, again in relation with increased activity of antioxidative enzymes (Yang et al., 2012). Furthermore, ammonium nutrition has been shown to increment rice tolerance to Fe deficiency through enhanced remobilization of Fe from root cell walls (Zhu et al., 2018).

CONCLUDING REMARKS AND FUTURE PERSPECTIVES

In this article, we propose a change of paradigm where ammonium nutrition may be considered not exclusively as an undesirable situation for plant performance, but as a way to provoke changes in plant metabolism that can be beneficial for crop quality and plant physiology. While some of the positive effects of ammonium referred here still require further evaluation, the cross-tolerance induction of NH₄⁺ to certain subsequent stresses, notably salinity, is clear. However, the molecular actors governing these interactions are almost completely unknown, and future works will be essential in order to fully exploit the benefits of ammonium-based fertilizers.

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DM and JM have made a substantial, direct, and intellectual contribution to the work and approved it for publication.

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REFERENCES

- Andrews, M., Condrón, L. M., Kemp, P. D., Topping, J. F., Lindsey, K., Hodge, S., et al. (2018). Elevated CO₂ effects on nitrogen assimilation and growth of C3 vascular plants are similar regardless of N-form assimilated. *J. Exp. Bot.* 70, 683–690. doi: 10.1093/jxb/ery371
- Bloom, A. J., Burger, M., Rubio-Asensio, J. S., and Cousins, A. B. (2010). Carbon dioxide enrichment inhibits nitrate assimilation in wheat and *Arabidopsis*. *Science* 328, 899–903. doi: 10.1126/science.1186440
- Bown, A. W., and Shelp, B. J. (2016). Plant GABA: not just a metabolite. *Trends Plant Sci.* 21, 811–813. doi: 10.1016/j.tplants.2016.08.001
- Bradáčová, K., Florea, A. S., Bar-Tal, A., Minz, D., Yermiyahu, U., Shawahna, R., et al. (2019). Microbial consortia versus single-strain inoculants: an advantage in PGPM-assisted tomato production? *Agronomy* 9, 105. doi: 10.3390/agronomy9020105
- Cao, X., Zhong, C., Zhu, C., Zhu, L., Zhang, J., Wu, L., et al. (2018). Ammonium uptake and metabolism alleviate PEG-induced water stress in rice seedlings. *Plant Physiol. Biochem.* 132, 128–137. doi: 10.1016/j.plaphy.2018.08.041
- Chaillou, S., Vessey, J. K., Morot-Gaudry, J. F., Raper, C. D., Henry, L. T., and Boutin, J. P. (1991). Expression of characteristics of ammonium nutrition as affected by pH of the root medium. *J. Exp. Bot.* 42, 189–196. doi: 10.1093/jxb/42.2.189
- Chi, W. J., Wang, Z. Y., Liu, J. M., Zhang, C., Wu, Y. H., and Bai, Y. J. (2019). Ammonium uptake and assimilation are required for rice defense against sheath blight disease. *Cereal Res. Commun.* 47, 98–110. doi: 10.1556/0806.46.2018.056
- Coletto, I., de la Peña, M., Rodríguez-Escalante, J., Bejarano, I., Glauser, G., Aparicio-Tejo, P. M., et al. (2017). Leaves play a central role in the adaptation of nitrogen and sulfur metabolism to ammonium nutrition in oilseed rape (*Brassica napus*). *BMC Plant Biol.* 17, 1–13. doi: 10.1186/s12870-017-1100-9
- de Souza Miranda, R., Mesquita, R. O., Costa, J. H., Alvarez-Pizarro, J. C., Prisco, J. T., and Gomes-Filho, E. (2017). Integrative control between proton pumps and SOS1 antiporters in roots is crucial for maintaining low Na⁺ accumulation and salt tolerance in ammonium-supplied *Sorghum bicolor*. *Plant Cell Physiol.* 58, 522–536. doi: 10.1093/pcp/pcw231
- Esteban, R., Ariz, I., Cruz, C., and Moran, J. F. (2016). Review: mechanisms of ammonium toxicity and the quest for tolerance. *Plant Sci.* 248, 92–101. doi: 10.1016/j.plantsci.2016.04.008
- Fernández-Crespo, E., Gómez-Pastor, R., Scalschi, L., Llorens, E., Camañes, G., and García-Agustín, P. (2014). NH₄⁺ induces antioxidant cellular machinery and provides resistance to salt stress in citrus plants. *Trees—Struct. Funct.* 28, 1693–1704. doi: 10.1007/s00468-014-1078-y
- Fernández-Crespo, E., Scalschi, L., Llorens, E., García-Agustín, P., and Camañes, G. (2015). NH₄⁺ protects tomato plants against *Pseudomonas syringae* by activation of systemic acquired acclimation. *J. Exp. Bot.* 66, 6777–6790. doi: 10.1093/jxb/erv382
- Fuertes-Mendizábal, T., González-Torralba, J., Arregui, L. M., González-Murua, C., González-Moro, M. B., and Estavillo, J. M. (2013). Ammonium as sole N source improves grain quality in wheat. *J. Sci. Food Agric.* 93, 2162–2171. doi: 10.1002/jsfa.6022

- Gao, Y., Li, Y., Yang, X., Li, H., Shen, Q., and Guo, S. (2010). Ammonium nutrition increases water absorption in rice seedlings (*Oryza sativa* L.) under water stress. *Plant Soil* 331, 193–201. doi: 10.1007/s11104-009-0245-1
- Gahoonia, T. S., Claassen, N., and Jungk, A. (1992). Mobilization of phosphate in different soils by ryegrass supplied with ammonium or nitrate. *Plant Soil* 140, 241–248. doi: 10.1007/BF00010600
- Gupta, K. J., Brotman, Y., Segu, S., Zeier, T., Zeier, J., Persijn, S. T., et al. (2013). The form of nitrogen nutrition affects resistance against *Pseudomonas syringae* pv. *phaseolicola* in tobacco. *J. Exp. Bot.* 64, 553–568. doi: 10.1093/jxb/ers348
- Hessini, K., Hamed, K. B., Gandour, M., Mejri, M., Abdely, C., and Cruz, C. (2013). Ammonium nutrition in the halophyte *Spartina alterniflora* under salt stress: evidence for a priming effect of ammonium? *Plant Soil* 370, 163–173. doi: 10.1007/s11104-013-1616-1
- Hessini, K., Kronzucker, H. J., Abdely, C., and Cruz, C. (2017). Drought stress obliterates the preference for ammonium as an N source in the C4 plant *Spartina alterniflora*. *J. Plant Physiol.* 213, 98–107. doi: 10.1016/j.jplph.2017.03.003
- Hessini, K., Issaoui, K., Ferchichi, S., Saif, T., Abdely, C., Siddique, K. H. M., et al. (2019). Interactive effects of salinity and nitrogen forms on plant growth, photosynthesis and osmotic adjustment in maize. *Plant Physiol. Biochem.* 139, 171–178. doi: 10.1016/j.plaphy.2019.03.005
- Huang, L., Li, M., Zhou, K., Sun, T., Hu, L., Li, C., et al. (2018). Uptake and metabolism of ammonium and nitrate in response to drought stress in *Malus prunifolia*. *Plant Physiol. Biochem.* 127, 185–193. doi: 10.1016/j.plaphy.2018.03.031
- Huber, D. M. (1989). "Introduction," in *Soilborne plant pathogens: management of diseases with macro-and microelements*. Ed. A. W. Engelhard (St Paul, Minn: APS Press), 1–8.
- Huber, H., and McCay-Buis, T. S. (1993). A multiple component analysis of the take-all disease of cereals. *Plant Dis.* 77, 437–447. doi: 10.1094/PD-77-0437
- Kinnersley, A. M., and Turano, F. J. (2000). Gamma aminobutyric acid (GABA) and plant responses to stress. *CRC Crit. Rev. Plant Sci.* 19, 479–509. doi: 10.1080/07352680091139277
- La, G.-X., Yang, T.-G., Fang, P., Guo, H.-X., Hao, X., and Huang, S.-M. (2013). Effect of $\text{NH}_4^+/\text{NO}_3^-$ ratios on the growth and bolting stem glucosinolate content of Chinese kale (*Brassica albuglabra* L.H. Bailey). *Aust. J. Crop Sci.* 7, 618–624.
- Leusch, H. J., and Buchenauer, H. (1988). Effect of soil treatments with silica-rich lime fertilizers and sodium trisilicate on the incidence of wheat by *Erysiphe graminis* and *Septoria nodorum* depending on the form of N-fertilizer. *J. Plant Dis. Prot.* 96, 154–172.
- Li, B., Li, G., Kronzucker, H. J., Baluška, F., and Shi, W. (2014). Ammonium stress in *Arabidopsis*: signaling, genetic loci, and physiological targets. *Trends Plant Sci.* 19, 107–114. doi: 10.1016/j.tplants.2013.09.004
- Liu, Y., and Von Wirén, N. (2017). Ammonium as a signal for physiological and morphological responses in plants. *J. Exp. Bot.* 68, 2581–2592. doi: 10.1093/jxb/erx086
- Logan, K. A. B., Thomas, R. J., and Raven, J. A. (2000). Effect of ammonium and phosphorus supply on h^+ production in gel by two tropical forage grasses. *J. Plant Nutr.* 23, 41–54. doi: 10.1080/01904160009381996
- Lopez-Berges, M. S., Rispail, N., Prados-Rosales, R. C., and Di Pietro, A. (2010). A nitrogen response pathway regulates virulence functions in *Fusarium oxysporum* via the protein kinase TOR and the bZIP protein MeaB. *Plant Cell* 22, 2459–2475. doi: 10.1105/tpc.110.075937
- Irigoyen, I., Lamsfus, C., Aparicio-Tejo, P., and Muro, J. (2006). The influence of 3,4-dimethylpyrazole phosphate and dicyandiamide on reducing nitrate accumulation in spinach under Mediterranean conditions. *J. Agric. Sci.* 144, 555–562. doi: 10.1017/S0021859606006472
- Marino, D., Ariz, I., Lasa, B., Santamaría, E., Fernández-Irigoyen, J., González-Murua, C., et al. (2016). Quantitative proteomics reveals the importance of nitrogen source to control glucosinolate metabolism in *Arabidopsis thaliana* and *Brassica oleracea*. *J. Exp. Bot.* 67, 3313–3323. doi: 10.1093/jxb/erw147
- Mpanga, I. K., Nkebiwe, P. M., Kuhlmann, M., Cozzolino, V., Piccolo, A., Geistlinger, J., et al. (2019). The form of N supply determines plant growth promotion by P-solubilizing microorganisms in maize. *Microorganisms* 7, 38. doi: 10.3390/microorganisms7020038
- Mur, L. A. J., Simpson, C., Kumari, A., Gupta, A. K., and Gupta, K. J. (2016). Moving nitrogen to the centre of plant defence against pathogens. *Ann. Bot.* 119, 703–709. doi: 10.1093/aob/mcw179
- Patterson, K., Cakmak, T., Cooper, A., Lager, I., Rasmusson, A. G., and Escobar, M. A. (2010). Distinct signalling pathways and transcriptome response signatures differentiate ammonium- and nitrate-supplied plants. *Plant Cell Environ.* 33, 1486–1501. doi: 10.1111/j.1365-3040.2010.02158.x
- Pastor, V., Gamir, J., Camaño, G., Cerezo, M., Sánchez-Bel, P., and Flors, V. (2014). Disruption of the ammonium transporter AMT1.1 alters basal defenses generating resistance against *Pseudomonas syringae* and *Plectosphaerella cucumerina*. *Front. Plant Sci.* 5, 1–16. doi: 10.3389/fpls.2014.00231
- Rubio-Asensio, J. S., and Bloom, A. J. (2017). Inorganic nitrogen form: a major player in wheat and *Arabidopsis* responses to elevated CO_2 . *J. Exp. Bot.* 68, 2611–2625. doi: 10.1093/jxb/erw465
- Sánchez-Rodríguez, E., Rubio-Wilhelmi, M. del M., Ríos, J. J., Blasco, B., Rosales, M. A., et al. (2011). Ammonia production and assimilation: its importance as a tolerance mechanism during moderate water deficit in tomato plants. *J. Plant Physiol.* 168, 816–823. doi: 10.1016/j.jplph.2010.11.018
- Santamaría, P., Gonnella, M., Elia, A., Parente, A., and Serio, F. (2001). Ways of reducing rocket salad nitrate content. *Acta Hort.* 548, 529–536. doi: 10.17660/ActaHortic.2001.548.64
- Santana, M. M., Gonzalez, J. M., and Cruz, C. (2017). Nitric oxide accumulation: the evolutionary trigger for phytopathogenesis. *Front. Microbiol.* 8, 1–13. doi: 10.3389/fmicb.2017.01947
- Sanz-Cobena, A., Lassaletta, L., Aguilera, E., del Prado, A., Garnier, J., Billen, G., et al. (2017). Strategies for greenhouse gas emissions mitigation in Mediterranean agriculture: a review. *Agric. Ecosyst. Environ.* 238, 5–24. doi: 10.1016/j.agee.2016.09.038
- Sarasketa, A., González-Moro, M. B., González-Murua, C., and Marino, D. (2014). Exploring ammonium tolerance in a large panel of *Arabidopsis thaliana* natural accessions. *J. Exp. Bot.* 65, 6023–6033. doi: 10.1093/jxb/eru342
- Sarasketa, A., González-Moro, M. B., González-Murua, C., and Marino, D. (2016). Nitrogen source and external medium pH interaction differentially affects root and shoot metabolism in *Arabidopsis*. *Front. Plant Sci.* 7, 29. doi: 10.3389/fpls.2016.00029
- Shavrukov, Y., and Hirai, Y. (2016). Good and bad protons: genetic aspects of acidity stress responses in plants. *J. Exp. Bot.* 67, 15–30. doi: 10.1093/jxb/erv437
- Traka, M. H., Saha, S., Huseby, S., Kopriva, S., Walley, P. G., Barker, G. C., et al. (2013). Genetic regulation of glucoraphanin accumulation in Beneforte broccoli. *New Phytol.* 198, 1085–1095. doi: 10.1111/nph.12232
- Umar, A. S., and Iqbal, M. (2007). Nitrate accumulation in plants, factors affecting the process and human health implications. A review. *Agron. Sustain. Dev.* 27, 45–57. doi: 10.1051/agro:2006021
- Vega-Mas, I., Marino, D., Sánchez-Zabala, J., González-Murua, C., Estavillo, J. M., and González-Moro, M. B. (2015). CO_2 enrichment modulates ammonium nutrition in tomato adjusting carbon and nitrogen metabolism to stomatal conductance. *Plant Sci.* 241, 32–44. doi: 10.1016/j.plantsci.2015.09.021
- Vega-Mas, I., Pérez-Delgado, C. M., Marino, D., Fuertes-Mendizábal, T., González-Murua, C., Márquez, A. J., et al. (2017). Elevated CO_2 induces root defensive mechanisms in tomato plants when dealing with ammonium toxicity. *Plant Cell Physiol.* 58, 2112–2125. doi: 10.1093/pcp/pcx146
- Wang, M., Sun, Y., Gu, Z., Wang, R., Sun, G., Zhu, C., et al. (2016). Nitrate protects cucumber plants against *Fusarium oxysporum* by regulating citrate exudation. *Plant Cell Physiol.* 57, 2001–2012. doi: 10.1093/pcp/pcw124
- Yang, X., Guo, S., Guo, L., and Wei, C. (2012). Ammonium enhances the uptake, bioaccumulation, and tolerance of phenanthrene in cucumber seedlings. *Plant Soil* 354, 185–195. doi: 10.1007/s11104-011-1054-x
- Zhu, C. Q., Zhang, J. H., Zhu, L. F., Abliz, B., Zhong, C., Bai, Z. G., et al. (2018). NH_4^+ facilitates iron reutilization in the cell walls of rice (*Oryza sativa*) roots under iron-deficiency conditions. *Environ. Exp. Bot.* 151, 21–31. doi: 10.1016/j.envexpbot.2018.03.018

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